



*8th
meeting on
medicinal
biotechnology*

BOOK OF
ABSTRACTS

MAY 29TH, 2026
SCHOOL OF HEALTH
POLYTECHNIC OF PORTO



BOOK OF ABSTRACTS OF THE 8TH MEETING ON MEDICINAL BIOTECHNOLOGY

School of Health - Polytechnic of Porto
Rua Dr. António Bernardino de Almeida, 400
4200 - 072, Porto
Portugal

ISBN: 978-989-9226-20-3
DOI: [HTTPS://DOI.ORG/10.26537/ED.P.PORTO.251](https://doi.org/10.26537/ED.P.PORTO.251)

May, 2026



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Impact of Simulated Digestion on Microplastics: Surface Changes and Intestinal *In Vitro* Exposure

MARIANA LAMAS^{1,2}, VIRGÍNIA CRUZ FERNANDES^{1,3}, MARTA OLIVEIRA¹ & FRANCISCA RODRIGUES^{1,5}

1. REQUIMTE/LAQV, ISEP, POLYTECHNIC OF PORTO, PORTO, PORTUGAL; 2. ICBAS - SCHOOL OF MEDICINE AND BIOMEDICAL SCIENCES, UNIVERSITY OF PORTO, PORTO, PORTUGAL; 3. RISE-HEALTH, CENTER FOR TRANSLATIONAL HEALTH AND MEDICAL BIOTECHNOLOGY RESEARCH (TBIO), ESS, POLYTECHNIC OF PORTO, PORTO, PORTUGAL; 4. I4HB - ASSOCIATE LABORATORY INSTITUTE FOR HEALTH AND BIOECONOMY, FACULTY OF PHARMACY, UNIVERSITY OF PORTO, PORTO, PORTUGAL; 5. UCIBIO - APPLIED MOLECULAR BIOSCIENCES UNIT, MEDTECH-LABORATORY OF PHARMACEUTICAL TECHNOLOGY, FACULTY OF PHARMACY, UNIVERSITY OF PORTO, PORTO, PORTUGAL.

Microplastics (MP) have been widely detected in a variety of food products¹, and ingestion is one of the main pathways for human exposure to these contaminants. Once ingested, MP might interact with the gastrointestinal (GI) environment, yet it remains unclear whether digestive processes alter their chemical structure or mainly influence their surface properties, and possibly their interaction capacity³. This study investigates digested-induced modifications in MP and establishes suitable conditions for intestinal *in vitro* assays. Simulated digestion of six polymers (PTFE; PMMA, PA6, uPVC, PET, and LDPE) followed the IN-FOGEST model². Following digestion, particles were analysed using Fourier-transform infrared spectroscopy (FTIR) to assess structural and surface changes. Cell viability was evaluated, by MTT, in Caco-2 and HT29-MTX intestinal cell lines exposed to different dilutions (1:10, 1:20, and 1:40) of digestion fluids, with

respective MP. FTIR results indicated that all polymers maintained their fundamental chemical structure after digestion. Instead, consistent surface-related alterations were observed, including reduced intensity of characteristic polymer bands and enhanced signals in regions associated with proteins and hydroxyl groups, suggesting biomolecules adsorption and increased hydration. Cell studies demonstrated that 1:20 dilution of digestive fluids with MP maintained the cell viability above 70%, representing the most suitable condition tested. Simulated GI digestion appears to preserve the intrinsic chemical identity of MP, while promoting surface modifications that may affect their biological interactions. The identification of 1:20 digestion fluid dilution supports its application in future studies using advanced intestinal models, including three-dimensional systems and assessment of cellular oxidative stress responses.

KEYWORDS: gastrointestinal digestion, chemical structure; intestinal cell models

FUNDING:

This work received financial support from the PT national funds (FCT/MECI) through the project UID/50006/2025 - Laboratório Associado para a Química Verde - Tecnologias e Processos Limpos and RISE-Health, R&D Unit (reference UID/06397/2025), and from the FEDER - the European Regional Development Fund (COMPETE 2030), through the WISE project - "Improving water for a safer environment" (No. 17091, ref. COMPETE2030-FEDER-00841800). M. Lamas (2024.05133.BD) would like to thank FCT/MECI and ESF for her PhD grant. M. Oliveira and F. Rodrigues thank to their TENURE programs (PRR No. 02/C06-i06/2024 and 2023.14718.TENURE.016, respectively).

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